

EVALUATION OF PHYTOCHEMICAL CONTENT, NUTRITIONAL VALUE AND ANTIOXIDANT ACTIVITY OF *OLAXSCANDENS* (ROXB) LEAVES

Raghavendra Naik¹, Sneha D Borkar², Acharya RN³, Shukla VJ⁴

¹PhD Scholar, Dravyaguna Department, ³Professor, Dravyaguna Department, ⁴Head- Pharmaceutical chemistry laboratory,IPGT&RA, Gujarat Ayurved University, Jamnagar, Gujarat, India.

²Assistant professor,Agadatantra Department, Mahatma JyotibaFule Ayurveda Mahavidyalaya, Chomu, Jaipur, India.

ABSTRACT

Leaves of *Olox scandens* (Roxb.) are consumed as vegetables in some parts of India. The present study was carried out to evaluate the phytochemical content, nutritional value and antioxidant activity of *Oloxscandens* (Roxb.) leaves following standard procedures. Qualitative analysis of the leaf powder reveals that carbohydrates, alkaloids, saponin, tannin and triterpenoids are present and glycosides, flavonoids, phenols and phytosterols are absent in leaves. The nutritional analysis of dried leaf powder of *O. scandens* shows the presence of macronutrients ie. Protein (12.89%w/w) carbohydrate (62.73%w/w) and fat (3.77%w/w) for each 100g dry sample. The leaf is also a good source for minerals like Calcium (2.52%), magnesium (0.77%) phosphorous (0.15 %) and Zinc (27.14mg/kg). Total antioxidant capacity of the leaf was 100.44±0.002 Mcg and IC50 values of the extract and Ascorbic acid were found to be >1000µg/ml and 11.67±0.58µg/ml respectively. Percentage scavenging of DPPH radical was found to rise with increasing concentration of the crude extract.

Key words: Antioxidants, Badru, DPPH, Leafy vegetables, Nutritional value, *Olox scandens* (Roxb.),

INTRODUCTION

Vegetables have been used as medicine since ancient times and playing a very important role in diet and nutrition. They are the most readily available sources of carbohydrates, fats, important proteins, vitamins, minerals, essential amino acids, and fibers¹. Their bioactive substances have a wide range of biological functions, including antioxidant and antimicrobial activities² and can be helpful in the management of oxidative stress and age related human ailments³. Regular consumption of fruits and vegetables has always been associated with health benefits, but their mechanism has become clear only in the recent decades. Fruits and vegetables contain a wide variety of biologically active, non-nutritive compounds known as phytochemicals. Leafy vegetables are natural source of antioxidants

and rich in phytochemicals^{4,5}. These phytochemicals impart health benefits beyond basic nutrition⁶. Fruits and vegetables contain different antioxidant compounds such as Vitamin C, vitamin E, and carotenoids, whose activities have been established in recent years. Flavonoids, tannins and other phenolic constituents present in food of plant origin are also potential antioxidants.⁷

Olox scandens (Roxb.), an ethno-medicinal plant, has been reported for its use as food and medicinal purpose. Different parts of the plant are used in conditions like fever, constipation, cough etc. Its leaves are roasted and eaten as vegetable⁸. Fresh young leaves chewed in mouth ulcer⁹. Fomentation of boiled leaves is applied externally in headache¹⁰. Further, its tender stem is also used as vegetable¹¹. Though

the plant *Oxaliscandens* is reported for many biological activities and used as vegetable, it is not evaluated for its chemical constituents and nutritional value. With the advent of modern systems of medicine need has been felt to investigate the active constituents present in these plants. Based on this, present study was undertaken to generate standardized data on various phyto and physico-chemical characteristics of leaves of the plant *Oxaliscandens* (Roxb.), along with nutritional composition and possible antioxidant activity. The outcome of the present study will be helpful to confirm its suitability as an edible vegetable.

MATERIALS AND METHODS:

Collection and preservation of the sample

Oxaliscandens (Roxb.), known as Badru, was identified from its natural habitat Balangir, Odisha, during September 2012, leaves were collected and authenticated by local taxonomist with the help of botanical flora.¹² A sample specimen was preserved in Pharmacognosy-

laboratory of IPGT & RA Jamnagar (SPECIMEN NO- PHM 6062/21/09/2012) and the sample was preserved in a solution prepared from 70% ethyl alcohol: glacial acetic acid: formalin (AAF) in the ratio of 90:5:5.¹³

Physico-chemical analysis

Physicochemical parameters and phytochemical screening were carried out as per the guidelines of Ayurvedic Pharmacopoeia of India.¹⁴

HPTLC study

Extraction of Alkaloids

Powdered plant materials were moistened with ammonium hydroxide and kept in a stoppered flask for about 1 hour. Then it was extracted with chloroform two-three times. About 5 ml chloroform extract was taken in a dish and chloroform evaporated. The dried substances were tested for the presence of alkaloid, and used for the chromatographic study. Chromatographic conditions are as follows.¹⁵

Chromatographic conditions

⇒ Application mode	:	CAMAG Linomate V
⇒ Development chamber	:	Camag Twin trough Chamber.
⇒ Plates	:	Pre-coated Silica Gel GF254 plates.
⇒ Chamber Saturation	:	30 min.
⇒ Development time	:	30 min.
⇒ Development distance	:	7 cm.
⇒ Scanner	:	Camag Scanner III.
⇒ Detection	:	Deuterium Lamp and Mercury Lamp.
⇒ Data system	:	Win cats software
⇒ Solvent system	:	Toluene: Ethyl acetate: formic acid
	7	: 2 : 0.5 v/v

Nutritional evaluation

Estimation of energy value- The sample caloric value was estimated (in Kcal) by multiplying the percentage crude protein, crude lipid and carbohydrate by the recommended factor (2.44, 8.37 and 3.57 respectively) used in analysis. The caloric value was determined based on the Atwater factor.¹⁶

Carbohydrates were determined by using cupric tartrate, the precipitate formed was com-

pared with dextrose of known concentration.¹⁷

Estimation of crude fat was performed using n – Hexane as solvent by Soxhlet extraction method.¹⁸ The crude protein was determined by the Kjeldahl method with slight modification and the absorbance at 470 nm.¹⁹ Determination of moisture content was carried out by standard procedure mentioned in Ayurvedic Pharmacopoeia of India.²⁰ All the minerals except phosphorus were analyzed from a triple

acid-digested sample by an atomic absorption spectrophotometer.²¹ The phosphorus content in the triple acid digested extract was determined colorimetrically.²²

Antioxidant assay

1,1-Diphenyl-2-picrylhydrazyl (DPPH) radical scavenging activity:

100 µM DPPH solution was obtained using methanol. 10.5 mg/ml, 10.5mg/ml and 21 mg/ml concentrations of ascorbic acid, rutin, and extract was obtained by using Dimethyl sulfoxide(DMSO) which was serially diluted with DMSO to obtain lower concentrations. Various concentrations of sample were added to DPPH solution and the absorbance of DPPH reagent was determined at 490 nm after 30 min of incubation, using a micro plate reader.²³

Total antioxidants assay

Weighed accurately 55 mg of the aqueous extract, standard, ascorbic acid and dissolved in

5 ml of DMSO. The lower dilutions were made serially with DMSO. An aliquot of 0.1 ml of the sample solution containing a reducing species in DMSO was combined with 1 ml of reagent solution (0.6 M Sulphuric acid, 28 mM sodium phosphate, and 4 mM ammonium molybdate). The tubes were capped and incubated in water bath at 95 °C for 90 min and the absorbance was measured at 695 nm. The total antioxidant capacity was expressed as mM equivalent of ascorbic acid.²⁴

RESULTS AND DISCUSSION

Powder of leaves was tested for various physico-chemical parameters such as loss on drying, ash value, water, methanol soluble extractive value. The observed results were depicted in table 1

Table 1: Preliminary Physico-chemical analysis of leaf powder of *Olax scandens* (Roxb.)

Sr. No.	Test	Results
1.	Loss on Drying	6.042 % w/w
2.	Ash Value	9.896 % w/w
3.	Acid Insoluble Ash	0.284 %w/w
4.	Water Soluble Extract	29.70% w/w
5.	Methanol Soluble Extract	22.40 % w/w
6.	pH	6.0

Preliminary qualitative chemical test:

Preliminary qualitative chemical test for leaves was done following standard procedure.

The observed results are presented in a tabular form (Table 2)

Table 2: Preliminary Qualitative analysis of leaf powder of *Olax scandens*.

S.No	Test	Results
1	Test for carbohydrates a. Molisch's test	+
2	Test for Glycosides a. Modified Borntrager's test b. Keller-Killiani test	- -
3	Test for Saponins a. Foam test	+
4	Test for Alkaloids a. Mayer's test b. Dragendorff's test	+ +

5	Test for Flavonoids a. Alkaline reagent test	-
6	Test for Phenolics and Tannins a. Ferric chloride test b. Test for Tannins	- +
	Test for Phytosterols and Triterpenoids a. Lieberman-Bucharat test b. Salkowaski test	- +
8	Test for fixed oils and fats a. Oily spot test	-

“+”: Positive, “-”: Negative

Carbohydrates, alkaloids, saponin, tannin and triterpenoids represent and Glycosides, flavonoids, phenols and phytosterols are absent in leaves.

HPTLC

Results of the HPTLC study of the leaves scanned under 254 nm & 366 nm showed 4 spots at 254 nm and 1 spot at 366 nm.

Table 3: Showing HPTLC profile for *Olax scandens*

Solvent system	Track No	Under UV light			
		254nm (Short UV)		366nm (Long UV)	
Toluene: Ethyl acetate: Formic acid 7.5: 2: 0.5 V/V	Track 1 (Leaves)	Number of spots	Rf value	Number of spots	Rf value
		4	0.16, 0.6, 0.66, 0.68	1	0.01

Nutritional analysis

The results of nutritional analysis of *Olax scandens* leaf is presented in table 4

Table 4: Nutritional values of leaf powder of *Olax scandens* (Roxb.)

Sr. No	Parameters	Results
1.	Energy	336.41 K Cals/100g
2.	Carbohydrate	62.73 gm
3.	Fat	3.77 gm
4.	Protein	12.89 gm
5.	Calcium	2.52 mg/Kg
6.	Magnesium	0.77 mg/Kg
7.	Phosphorous	0.15 mg/Kg
8.	Zinc	27.14 mg/Kg

The results obtained in the nutritional analysis of dried leaf powder of *O. scandens* shows the presence of macronutrients ie. protein (12.89%w/w) carbohydrate (62.73%w/w) and fat (3.77%w/w) for each 100g dry sample. Macronutrients are nutrients that provide calories or energy. These are substances needed

for growth, metabolism, and for other body functions. According to the Dietary Reference Intakes published by the USDA, 45% - 65% of calories should come from carbohydrate, 10% - 35% of calories should come from protein and 20% - 35% of calories should come from fat.

The leaf is also a good source for minerals like Calcium (2.52%), magnesium (0.77%) phosphorous (0.15 %) and Zinc (27.14mg/kg).

Calcium is an essential mineral for bone formation, deficiency of which leads to reduced bone formation, osteoporosis and bone fracture with an overall restriction in bone formation (bone size) and growth. Magnesium plays a vital role in the activity of many enzymes and phosphorous is an important component of energy intermediates²⁵.

Leaves showed higher values of Zinc (27.14 mg/Kg) indicating them as a good source of this vital trace element. Zinc is a component of many metallo enzymes and also a membrane stabilizer and a stimulator of the immune response.^{26,27} Its deficiency leads to loss of appetite, and impaired immune function. In more severe cases, zinc deficiency causes hair loss, diarrhea, delayed sexual maturation, impotence, hypogonadism in males, and eye and skin lesions.^{28,29} The high zinc content may be useful in skin diseases and rheumatism.

The value of zinc is found to be higher than that of the previous study.³⁰ The variability in

the content in the same species may be related to genetic origin, geographical source and the levels of soil fertility. The present result indicates the potentiality of plant *Olařscandens* as source of unconventional food. Being wild, it is easily accessible and cheaper vegetable source.

Antioxidant activity

DPPH radical scavenging activity

Results of the DPPH radical scavenging assay is given in Table 5. The IC₅₀ values of the extract and Ascorbic acid were found to be >1000µg/ml and 11.67±0.58µg/ml respectively. Percentage scavenging of DPPH radical was found to rise with increasing concentration of the crude extract (Figure 1).

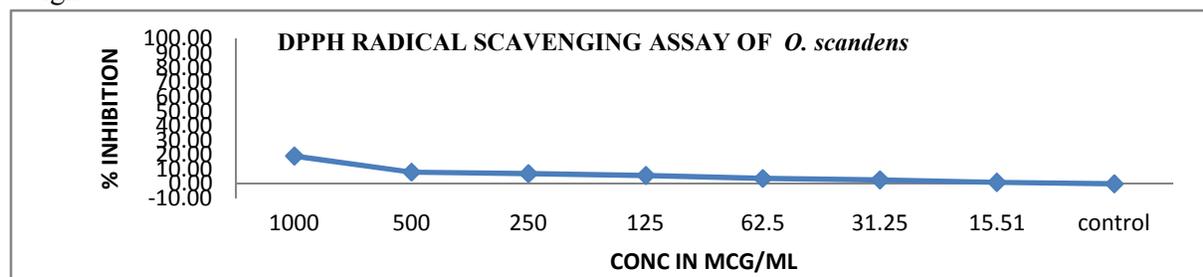
Total antioxidant capacity

Total antioxidant capacity of the extract is 100.44±0.002 Mcg (Table 5). Total antioxidant capacity is expressed as the number of equivalents of ascorbic acid (AAE).

Table 5: DPPH radical scavenging activity and total antioxidant capacity of *O. scandens*

Samples	IC ₅₀ values µg/ml by methods	Mcg per equivalent of Ascorbic acid
	DPPH	Total antioxidant activity* ^a
<i>O.scandens</i>	>1000	100.44±0.002
Standard	Ascorbic acid	
	11.67±0.58	

*^aThe total antioxidant capacity was expressed as mg equivalent of ascorbic acid per gram of dry weight.



CONCLUSION

Observed physicochemical and phytochemical parameters can be considered as the standard

for future references. Leaves of *O. scandens* area good source of macronutrients ie. Protein, carbohydrate and fat and minerals like

calcium, magnesium, phosphorous and zinc. Leaves of *O. scandens* also possess mild antioxidant activity when compared with the ascorbic acid. The results of present study indicates the potentiality of plant *Olaixscandens* as

source of unconventional food. Being wild, it is easily accessible and cheaper vegetable source.

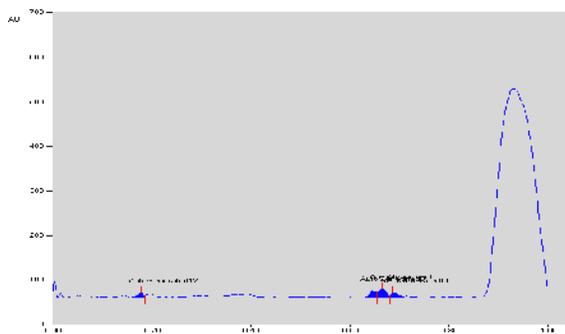
PHOTOGRAPHS PLATE A



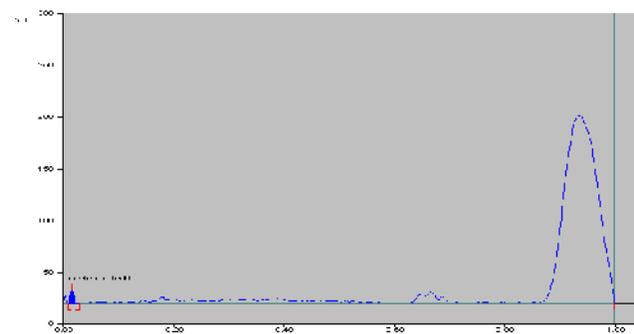
TLC Plate at 254 nm



TLC Plate at 366 nm



Peak Display at 254 nm



Peak Display at 366nm

REFERENCES

1. Sharma HP, Kumar RA. Health security in ethnic communities through nutraceutical leafy vegetables. *J Environ Res Develop.* 2013;7(4):1423–1429
2. Kim SJ, Cho AR, Han J. Antioxidant and antimicrobial activities of leafy green vegetable extracts and their applications to meat product preservation. *Food Control.* 2013;29:112–120.
3. Gacch RN, Kabaliye VN, Dhole NA, Jadhav AD. Antioxidant potential of selected vegetables commonly used in diet in Asian subcontinent. *Indian J Nat Prod Resour.* 2010;1(3):306–313.
4. Elias KM, Nelson KO, Simon M, Johnson K. Phytochemical and antioxidant analysis of methanolic extracts of four African indigenous leafy vegetables. *Ann Food Sci Technol.* 2012;13(1):37–42
5. Raghavendra M, Reddy AM, Yadav PR, Raju AS, Kumar LS. Comparative studies on the in vitro antioxidant properties of methanolic leafy extracts from six edible leafy vegetables of india. *Asian J Pharm Clin Res.* 2013;6(3):96–99.
6. Oomah BD, Mazza G (2000) Functional foods. In: Francis FJ (ed) *The Wiley encyclopedia of science & technology*, 2nd edn. Wiley, New York, pp 1176–1182
7. Salah N, Miller NJ, Paganga G, Tijburg L, Bolwell GP and RiceE-vansCl. Polyphenolic flavanols as

- scavengers of aqueous phase radicals and as chain breaking antioxidants. Archives of Biochemistry and Biophysics 1995;322(2):339-346.
8. Sibangini Misra and Malaya K. Misra, Leafy Vegetable Plants of South Odisha, India, International Journal of Agricultural and Food Science, 2013, 3(4): 131-137.
 9. <https://sites.google.com/site/efloraofindia/species/m.../olax/olax-scandens>
 10. Anonymous, (1990), Glimpses of medicobotany of Bastar district, Madhya Pradesh, CCRAS publication, pp. 114.
 11. Sibangini Misra and Malaya K. Misra, Leafy Vegetable Plants of South Odisha, India, International Journal of Agricultural and Food Science, 2013, 3(4): 131-137.
 12. Saxena H.O, (1995), The Flora of Orissa, Regional Research Laboratory, Bhubaneswar, 1st edition, pp. 288.
 13. Johnson Alexander Donald, (1940), Plant Micro techniques, Macgrew Hill Book Company, New York, London. pp. 105.
 14. Anonymous, The Ayurvedic Pharmacopoeia of India, 1st ed, Govt. of India. Ministry of Health and Family welfare, Department of I.S.M. & H., New Delhi 1999; I: Appendix 2.
 15. Anonymous, Planner Chromatography, Modern Thin layer Chromatography, Switzerland (1999), pg. 2-16
 16. FAO Corporate Document Repository. Agriculture and consumer protection: calculation of the energy content of foods - energy conversion factors (2006).[cited 2014 April 25]; Available from: <http://www.fao.org/docrep/006/y5022e/y5022e04.htm>
 17. Anonymous Association of Official Analytical Chemists. Official Methods of Analysis of the AOAC. 15th ed. Washington: D.C.; 1990. pp. 375 – 379.
 18. Anonymous Association of Official Analytical Chemists. Official Methods of Analysis of the AOAC. 15th ed. Washington: D.C.; 1990. pp. 375 – 379.
 19. Anonymous Association of Official Analytical Chemists. Official Methods of Analysis of the AOAC. 15th ed. Washington: D.C.; 1990. pp. 375 – 379.
 20. Anonymous. The Ayurvedic Pharmacopoeia of India, 1st ed. New Delhi: Govt. of India, Ministry of Health and Family welfare, Department of I.S.M. & H.; 1999. Appendix 2 (2.2.3). pp. 213
 21. Issac, R.A., Johnson, W.C. Collaborative study of wet and dry techniques for the elemental analysis of plant tissue by Atomic Absorption Spectrophotometer. J Ass Off Analy Chem 1975; 58:436-440.
 22. ¹Dickman, S.R., Bray, R.H. Colorimetric determination of phosphate. Ind Eng Chem Anal 1940; 12:665 – 668.
 23. VK Jinesh, V Jaishree, Shrishailappa Badami and W Shyam. Comparative evaluation of antioxidant properties of edible and non-edible leaves of *Anethum graveolens* Linn. Indian Journal of Natural Products and Resources 2010; 1(2):168-173.
 24. Anonymous Association of Official Analytical Chemists. Official Methods of Analysis of the AOAC. 15th ed. Washington: D.C.; 1990. pp. 375 – 379.
 25. Vance, C. P., Uhde-Stone, C. & Allan, D. L. (2003). Phosphorus acquisition and use: critical adaptations by plants for securing a nonrenewable resource. *New Phytologist* 157: 432-449
 26. Hambidge KM. Zinc as membrane stabilizer. J Hum Nut. 1978;32:99-100.
 27. Solomons NW. Mild human zinc deficiency produces an imbalance between cell-mediated and humoral immunity. Nutr Rev. 1998;56:27-8.
 28. Maret W, Sandstead HH. Zinc requirements and the risks and benefits of zinc

- supplementation. J Trace Elem Med Biol.2006;20:3–18.
29. Prasad AS. Zinc deficiency: Its characterization and treatment. Met Ions Biol Syst. 2004;41:103–37.
30. Sunita Thakur, Sudhanshu Kumar, Arvind Kumar, Potential Of Some Wild Leafy Vegetables As Natural Source For Supplementation Of Micronutrients In Vegetarian Diets Of SanthalPargana Area Of Jharkhand, Journal of Fundamental and

Applied Life Sciences 2012 Vol. 2 (3) July-September, pp.65-67

CORRESPONDING AUTHOR

Dr. Raghavendra Naik

PhD Scholar, Department of Dravyaguna, IPGT&RA, Gujarat Ayurved University, Jamnagar, Gujarat, India

Email: ayuraghu@gmail.com

Source of support: Nil

Conflict of interest: None Declared